

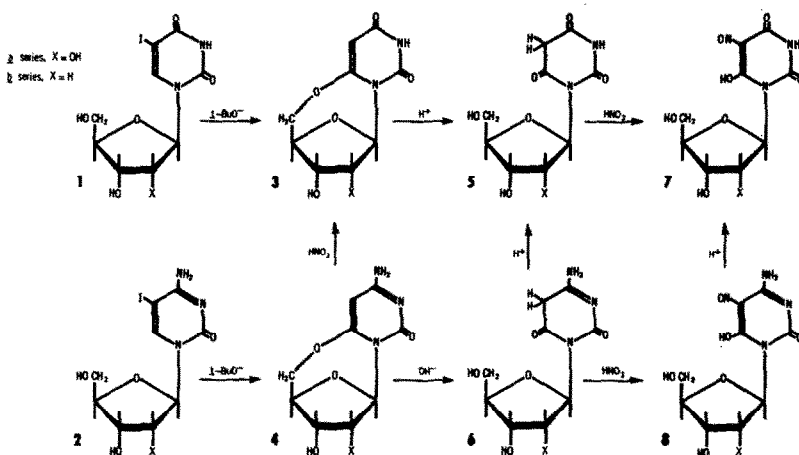
0<sup>6</sup>,5'-CYCLONUCLEOSIDES. REACTIONS OF 5-IODOPYRIMIDINE NUCLEOSIDES WITH BASE<sup>(1)</sup>

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The recent note by Fox *et al.*,<sup>(3)</sup> prompts us to report the synthesis of four 0<sup>6</sup>,5'-cyclopyrimidine nucleosides<sup>(4)</sup> (3a, 3b, 4a, 4b) from the corresponding 5-iodopyrimidine nucleosides.<sup>(5)</sup> Such cyclonucleosides are of interest since they can be used as intermediates in the synthesis of 6-substituted pyrimidine nucleosides, a class of compounds that may prove to be of biological importance, but which to date have been little explored.<sup>(6,7,8)</sup> Thus, when 5-iodouridine (1a) was treated with an excess of potassium *t*-butoxide in a 1:1 (v/v) mixture of *t*-butyl alcohol and DMSO (60°, 24 hr), 0<sup>6</sup>,5'-cycloouridine<sup>(9)</sup> (3a) [mp 283-285° dec;  $\lambda_{\text{max}}^{\text{pH } 7}$  262 m $\mu$  ( $\epsilon$ , 12,080)] was obtained. Analogously, starting with 5-iodocytidine (2a) and 5-iodo-2'-deoxycytidine (2b), 0<sup>6</sup>,5'-cyclocytidine·1/2H<sub>2</sub>O (4a) [mp > 320°,  $\lambda_{\text{max}}^{\text{pH } 7}$  271.5 m $\mu$  ( $\epsilon$ , 11,320)] and 0<sup>6</sup>,5'-cyclo-2'-deoxycytidine (4b) [mp > 300°,  $\lambda_{\text{max}}^{\text{pH } 7}$  272 m $\mu$  ( $\epsilon$ , 10,400)], resp., were obtained. When 5-iodo-2'-



deoxyuridine (1b) replaced 1a in the above type of reaction, only a 19% yield of 0<sup>6</sup>,5'-cyclo-2'-deoxyuridine·1/2H<sub>2</sub>O<sup>(10)</sup> (3b) [mp 209-210° dec;  $\lambda_{\text{max}}^{\text{pH } 7}$  262.5 m $\mu$  ( $\epsilon$ , 12,600)] was isolated. The remainder of the reaction mixture was 40% unreacted 1b, 2% barbituric acid (13), and 27% 6-hydroxy-2'-deoxyuridine (5b). Preparatively it proved advantageous to obtain 3b by treating 4b with an excess of LiNO<sub>2</sub> and glacial acetic acid in DMSO.

The structures of 3a and 3b were confirmed by the following observations. Treatment of 3a and 3b with 60% HF<sup>(5,11)</sup> (25°, 15 hr) yielded 13 and ribose (14) or deoxyribose (15), resp. On the other hand, 0.1N H<sub>2</sub>SO<sub>4</sub> converted 3a and 3b (70°, 1 hr) to 6-hydroxyuridine·H<sub>2</sub>O (5a) [mp 114.5–115°;  $\lambda_{\max}^{\text{pH } 7}$  264 m $\mu$  ( $\epsilon$ , 21,890)] and 6-hydroxy-2'-deoxyuridine (5b) ( $\lambda_{\max}^{\text{pH } 7}$  265 m $\mu$ ), resp. This indicates that the position of attachment of the cyclo-linkage is at C<sub>6</sub> of the aglycon. It is possible to differentiate 5a from the isomeric 5-hydroxyuridine<sup>(12)</sup> (16) [ $\lambda_{\max}^{\text{pH } 7}$  280 m $\mu$  ( $\epsilon$ , 8,200)] by comparison of their UV spectra; the lack of blue color when 5a is treated with aqueous FeCl<sub>3</sub>,<sup>(13,14)</sup> and the orange color produced when 5a or 13 are treated with Ehrlich's reagent.<sup>(15)</sup> No color is produced when 16 is treated with Ehrlich's reagent. NMR spectra of 3a and 3b in DMSO-d<sub>6</sub> showed a single vinylic proton at 5.32  $\delta$  and 5.19  $\delta$ , resp. These singlet resonances are assigned to the H<sub>5</sub> protons<sup>(16a)</sup> of 3a and 3b. The H<sub>5'</sub> resonances of 3a and 3b, on the other hand, are pairs of doublets<sup>(16b)</sup> centered at 3.95  $\delta$  and 4.58  $\delta$  ( $J_{\text{H}_5', \text{H}_5} = 13$  cps) and 3.89  $\delta$  and 4.51  $\delta$  ( $J_{\text{H}_5', \text{H}_5} = 12$  cps), resp. Such a pattern has been reported<sup>(3,17)</sup> as typical of cyclonucleosides that have an oxygen cyclo-linkage to C<sub>5'</sub>. Compound 3a rapidly consumed one mole of metaperiodate.<sup>(18)</sup> This is consistent with a ribofuranosyl structure and a cyclo-linkage which does not involve C<sub>2'</sub> or C<sub>3'</sub>.

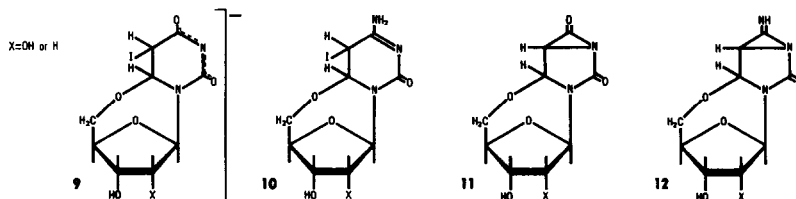
Similar methods were used to determine the structures of 4a and 4b. Using deaminating conditions analogous to those used for the conversion of 4b to 3b, it was possible to convert 4a to 3a. This indicates that the cyclo-linkage does not involve C<sub>4</sub> of the aglycon. It was possible, furthermore, to open the cyclo-linkage of 4a and 4b by refluxing (1 hr) in aqueous 0.2M Ba(OH)<sub>2</sub>. 6-Hydroxycytidine·H<sub>2</sub>O (6a) [mp 181–182° dec;  $\lambda_{\max}^{\text{pH } 7}$  267 m $\mu$  ( $\epsilon$ , 19,800)] and 6-hydroxy-2'-deoxycytidine·H<sub>2</sub>O (6b) [mp 195–197° dec;  $\lambda_{\max}^{\text{pH } 7}$  267 m $\mu$  ( $\epsilon$ , 19,990)], resp., were obtained. The H<sub>5</sub> protons<sup>(16a)</sup> of 4a and 4b were single unsplit resonances at 5.40  $\delta$  and 5.33  $\delta$  resp., while the H<sub>5'</sub> protons in each were again pairs of doublets centered at 4.50  $\delta$  and 3.87  $\delta$  ( $J_{\text{H}_5', \text{H}_5} = 14$  cps) and 4.49  $\delta$  and 3.82  $\delta$  ( $J_{\text{H}_5', \text{H}_5} = 13$  cps), resp. Compound 4a rapidly consumed one mole of metaperiodate. Once again, this is consistent with a ribofuranosyl structure and a cyclo-linkage that is to C<sub>5'</sub> rather than to C<sub>2'</sub> or C<sub>3'</sub>.

By treatment of 2a and 2b with excess aqueous tetramethylammonium hydroxide in DMSO (25°, 3 days) it is possible to isolate 6a and 6b, resp. In all likelihood 4a and 4b, resp., are intermediates. This supposition is supported by the observations that during the conversion of 2a to 6a a small amount of 4a always is present and that the cyclo-linkage in 4a and 4b is opened with Ba(OH)<sub>2</sub> to form 6a and 6b, resp. Compounds 6a and 6b can be deaminated to 5a and 5b, resp.,

by treatment with aqueous 0.1M H<sub>2</sub>SO<sub>4</sub> (25°, 18 hr). The structures of 6a and 6b were further confirmed by cleavage with 60% HF (25°, 16 hr) to 6-hydroxycytosine (17) and 14 or 15, resp. It was possible also to distinguish 6a from the isomeric 5-hydroxycytidine<sup>(19)</sup> [ $\lambda_{\text{max}}^{\text{pH } 7}$  292 m $\mu$  ( $\epsilon$ , 7837)] by comparison of their UV spectra and the lack of blue color when 6a was treated with aqueous FeCl<sub>3</sub>.<sup>(13)</sup> Finally, it was possible to convert 5a to 5-nitroso-6-hydroxyuridine (7a) [mp 166-167° dec;  $\lambda_{\text{max}}^{\text{pH } 7}$  311 m $\mu$  ( $\epsilon$ , 8000)] by treatment with excess HNO<sub>2</sub>. This is further evidence that the cyclo-linkage in 3a does not involve C<sub>5</sub> of the aglycon. The structure of 7a was confirmed by comparison of its UV spectrum with violuric acid [ $\lambda_{\text{max}}^{\text{pH } 7}$  311 m $\mu$ ]. Likewise, reaction of 6a with aqueous HNO<sub>2</sub> at pH > 4 yielded 5-nitroso-6-hydroxycytidine·1/2H<sub>2</sub>O (8a) [mp 210-211° dec;  $\lambda_{\text{max}}^{\text{pH } 7}$  317 m $\mu$  ( $\epsilon$ , 17,400)]. Compound 8a then was deaminated to 7a by treatment with aqueous 0.1M H<sub>2</sub>SO<sub>4</sub> (25°, 15 hr).

Two plausible mechanisms can explain the conversion of 5-iodopyrimidine nucleosides to the corresponding 0<sup>6</sup>,5'-cyclopyrimidine nucleosides. The first is an addition-elimination mechanism. This would involve the anion 9 or the intermediate 10, which could either eliminate HI directly by the loss of H<sub>6</sub> and iodide<sup>(3)</sup> or it could form the  $\alpha$ -lactam 11 or the  $\alpha$ -iminolactam 12 by displacement of iodide at C<sub>5</sub> by N<sub>3</sub>. H<sub>6</sub> could then be lost with concomitant lactam ring opening to form the observed product. Related mechanisms<sup>(20,21,22)</sup> have been proposed to explain reaction at C<sub>6</sub> in various other pyrimidine nucleosides and the conversion<sup>(23)</sup> of showdomycin to cycloshowdomycin. The second plausible mechanism involves a hetaryne. In this case the first step is abstraction of H<sub>6</sub> by base followed by loss of iodide and the attack of the C<sub>5</sub>' oxygen on the hetaryne. Hetaryne intermediates have been observed with other 5-halopyrimidines.<sup>(24)</sup> Evidence obtained thus far in This Laboratory supports either mechanism. No matter which is operative, however, it need not function only intramolecularly, since treatment of 5-iodocytidine-5'-phosphate<sup>(25)</sup> with aqueous tetramethylammonium hydroxide in DMSO yields the 5'-phosphate of 4a.

Further work is in progress in order to elucidate mechanisms for these reactions and to determine the scope of ring-opening reactions of the cyclonucleosides with common nucleophiles.



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